Prostatic Intraepithelial Neoplasia (PIN): Current Concepts

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Abstract Prostatic intraepithelial neoplasia (PIN) represents the putative precancerous end of the morphologic continuum of cellular proliferations within prostatic ducts, ductules and acini. Two grades of PIN are identified (low grade and high grade), and high grade PIN is considered to be a precursor to invasive carcinoma. The continuum which culminates in high grade PIN and early invasive cancer is characterized by basal cell layer disruption, basement membrane disruption, progressive loss of secretory differentiation markers, increasing nuclear and nucleolar abnormalities, increasing proliferative potential, and increasing variation in DNA content (aneuploidy). Clinical studies suggest that PIN predates carcinoma by ten years or more, with low grade PIN first emerging in men in the third decade of life. The clinical importance of recognizing PIN is based on its strong association with carcinoma; its identification in biopsy specimens of the prostate warrants further search for concurrent invasive carcinoma.

Key words: cancer, dysplasia, hyperplasia, prostate, prostatic intraepithelial neoplasia (PIN)

Despite the potential importance of a premalignant phase in the natural history of human cancer, little attention has been given to the study of precursor lesions of malignancy in the prostate. Recently, two potential morphologic precursors of adenocarcinoma have been identified, referred to as prostatic intraepithelial neoplasia (PIN) and atypical adenomatous hyperplasia (AAH). These lesions display some of the features of carcinoma, but PIN lacks stromal invasiveness and AAH lacks significant cytologic atypia. High grade PIN is commonly found in association with invasive cancer, and the high predictive value of this lesion suggests that further search for carcinoma be made in cases in which high grade PIN is found on biopsy. AAH is also found in association with carcinoma, but not as commonly as PIN. In 1991, an international consensus conference group sponsored by the American Cancer Society concluded that PIN was the most likely precursor of invasive carcinoma, and that AAH should be further investigated as a possible precursor.

This report reviews the diagnostic criteria and clinical significance of PIN, including the evidence accumulated to date linking this histopathologic lesion with invasive cancer.

DIAGNOSTIC CRITERIA FOR PIN

PIN refers to the putative precancerous (dysplastic) end of the morphologic continuum of cellular proliferations within prostatic ducts, ductules, and acini [1–6]. The term PIN was endorsed by consensus at a 1989 international conference [1–7] to replace other synonymous terms used in the literature, including intraductal dysplasia [8], large acinar atypical hyperplasia [9], atypical primary hyperplasia [10], hyperplasia with malignant change [11], marked atypia [12], and duct-acinar dysplasia [13-15]. This consensus group also agreed that PIN should be divided into two grades (low grade and high grade) to replace the previous three grade system (PIN 1 is considered low grade, and PIN 2 and 3 are considered high grade) [7].

In low grade PIN, the cells within ducts and acini are heaped up, crowded, and irregularly spaced with marked variation in nuclear size (anisonucleosis) (Table 1; Fig. 1). Elongate hyperchromatic nuclei and small nucleoli are also observed, but these are not usually prominent features. The diagnosis of PIN requires a combination of both cytologic and architectural features, and lesions displaying some but not all of these features are considered atypical but not

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TABLE 1. PROSTATIC INTRAEPITHELIAL NEOPLASIA (PIN): DIAGNOSTIC CRITERIA

	Low Grade PIN (Formerty PIN 1)	High Grade PiN (Formerty PIN 2 and 3)
ARCHITECTURE	Epithelial cell crowding and stratification, with irregular spacing.	Similar to low grade PIN; more crowding and stratification; 4 patterns: tufting, micropapillary, cribriform, and flat.
CYTOLOGY		
Nuclei	Enlarged, with marked size variation	Enlarged; some size and shape variation
Chromatin	Normal	increased density and clumping
Nucleoli	Rarely prominent	Occasionally to frequently large and prominent, similar to invasive carcinoma; sometimes multiple
BASALCELL LAYER	intact	May show some disruption
BASEMENT MEMBRANE	intact	May show some disruption

Modified from Bostwick and Brawer [2].

dysplastic. High grade PIN (formerly PIN 2 and 3) exhibits features similar to low grade PIN, although cell crowding and stratification are usually more pronounced, with less variability in nuclear size because the majority of nuclei are enlarged; the presence of prominent nucleoli, often numerous, is of greatest diagnostic utility (Fig. 2). We have identified four patterns of high grade PIN: tufting, micropapillary, cribriform, and flat [16].

PIN spreads through prostatic ducts in three different patterns, similar to prostatic carcinoma [5,17]. In the first pattern, neoplastic cells replace the normal luminal secretory epithelium, with preservation of the basal cell layer and basement membrane. Foci of high grade PIN are usually indistinguishable from ductal spread of carcinoma by routine light microscopy in our experience, although some authors claim that this distinction is possible [18]. In the second pattern, there is direct invasion through the ductal or acinar wall, with disruption of the basal cell layer and the basement membrane. In the third pattern, neoplastic cells invaginate between the basal cell layer and columnar secretory cell layer ("pagetoid spread"), a very uncommon finding.

PIN AND CANCER OCCUR COMMONLY IN THE PERIPHERAL ZONE

The peripheral zone of the prostate, the area in which the majority of prostatic carcinomas occur (70%), is also the most common location for PIN [19]. Cancer and PIN are frequently multifocal in the peripheral zone, indicating a "field" effect similar to the multi-focality of transitional cell carcinoma of the bladder. The transition zone and periurethral area, the anatomic areas in which nodular hyperplasia occurs, account for about 20–25% of prostate cancers and harbor foci of PIN in only 2% of cases [20]. By contrast, AAH is found in up to 24% of transition zone specimens [20–22]. If AAH is not a precursor of carcinoma, then the infrequency of PIN in the transition zone could account for the infrequency of cancer originating in that area.

THE BASAL CELL LAYER AND BASEMENT MEMBRANE ARE DISRUPTED IN HIGH GRADE PIN

Increasing grades of PIN are associated with progressive disruption of the basal cell layer and basement membrane [2,23-25]. Basal cell-specific monoclonal antibodies directed against high molecular weight keratin (e.g., clone 34β -E12) have been employed immunohistochemically to selectively label the prostatic basal cell layer [2]. Tumor cells consistently failed to be decorated with this antibody, whereas normal prostatic epithelium was invariably stained, with a continuous intact circumferential basal cell laver observed in most instances. Basal cell layer disruption was present in 56% of cases of high grade PIN, and more commonly in glands adjacent to invasive carcinoma than in distant glands. Also, the amount of disruption increased with increasing grades of PIN, with loss of more than one-third of the basal cell layer in 52% of foci of high grade PIN [2].

The type IV collagen-immunoreactive basement membrane normally surrounding prostatic glands was focally attenuated or absent in 40% of cases of PIN [2–3]. Preliminary data have shown increased expression of type IV collagenase in PIN and cancer [26]; collagenase is a proteolytic enzyme which is thought to induce fragmentation of the basement membrane during invasion [27].

Early invasive carcinoma occurs at sites of glandular out-pouching and basal cell disruption [2,15]. Although one author has referred to this as "transitive" gland change [15], we feel that "microinvasion" is the preferred terminology as it is in other organs, avoiding introduction of a new and unnecessary term. A model of

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Fig. 1. Low grade PIN. The cells are crowded and irregularly spaced, with variation in nuclear size. Nucleoli are occasionally observed, but are not prominent.



Fig. 2. High grade PIN. There is cell crowding and stratification, with nuclear enlargement and nucleomegaly. Cytoplasmic apical blebs are noted in more than 90% of cases of high grade PIN.

Prostatic Intraepithelial Neoplasia



Fig. 3. Morphologic continuum from normal prostatic epithelium through increasing grades of PIN to early invasive carcinoma, according to the disease-continuum concept. Low grade PIN (grade 1) corresponds to very mild to mild dysplasia. High grade PIN (grades 2 and 3) corresponds to moderate to severe dysplasia and carcinoma *in situ*. The precursor state ends when malignant cells invade the stroma; this invasion occurs where the basal

prostatic carcinogenesis has been proposed based on the morphologic continuum of PIN and the multi-step theory of transformation (Fig. 3) [2].

THERE IS INCREASED FREQUENCY, SEVERITY, AND EXTENT OF PIN WITH CANCER

The frequency of PIN in prostates with cancer was significantly higher when compared with prostates without cancer (Fig. 4) [8–9, 28–32]. We observed PIN in 82% of step-sectioned prostates with cancer, but in only 43% of benign prostates from patients of similar age [8]. PIN was more extensive in amount in lower stage tumors, presumably due to "overgrowth" or obliteration of PIN in prostates with larger high-stage tumors [15,33].

The severity of PIN in prostates with cancer was significantly higher when compared with prostates without cancer [8].

PIN AND CANCER INCREASE WITH PATIENT AGE

In a study of 429 step-sectioned whole prostates, Kovi *et al.* found that the prevalence of PIN in prostates with cancer increased with age, predating the onset of carcinoma by more than five years [9]. A similar study by Sakr *et al.* revealed the presence of PIN in men in their cell layer is disrupted and the basement membrane is fragmented. Notice that the dysplastic changes occur in the superficial (luminal) secretory cell layer, perhaps in response to luminal carcinogens. Disruption of the basal cell layer and basement membrane accompanies the architectural and cytologic features of high grade PIN, and appears to be a necessary prerequisite for stromal invasion.



Fig. 4. Frequency of PIN and AAH in autopsy prostates. These data are from an autopsy series of 100 serially sectioned prostates with cancer and 100 prostates without cancer [8]. Note that the frequency of both of these putative premalignant lesions (PIN and AAH) is significantly increased in prostates with cancer when compared with those not harboring cancer, although PIN is significantly more common.

twenties and thirties (9% and 22% frequency, respectively), which preceded the onset of carcinoma by more than ten years (Fig. 5) [34]. Most foci of PIN in young males were low grade, with increasing frequency of high grade PIN with advancing age. The prevalence of PIN was similar in blacks and whites.

Lee *et al.* studied 256 ultrasound-guided biopsies of hypoechoic lesions of the prostate, and identified 103 cancers and 27 cases of PIN;

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TABLE 2. PHENOTYPE OF PROSTATIC INTRAEPITHELIAL NEOPLASIA (PIN)

	Expression in PIN Compared with Normal Epithelium	Expression in PIN Compared with Cancer	References
Cytoplasmic Proteins			
Prostate Specific Antigen (PSA)	Decreased	Variable	14
Prostatic Acid Phosphatase (PAP)	Decreased	Variable	14
Leu-7	Decreased	Variable	14
Pepsinogen II (PG II)	Decreased	Decreased	15
Tissue Plasminonen Activator (t-PA)	Elevated	Elevated	15
Type IV Collagenase	Elevated	Same	26
Intermediate Filaments			
Cytokeratins 14,15,16,19	Elevated	Same	37
Vimentin	Decreased	Same	37
Lectins			
Ulex Europaeus (UEA-1)	Elevated	Same	37, 38
	Decreased	Same	14, 39
Concavalia ensilormis (Con-A)	Decreased	Same	14, 39
Arachis Hypogen (PNA)	Decreased	Same	14, 39
Bandeirea simplificifolia (BS-I)	Decreased	Not evaluated	14
Dolichos biflorus (DBA)	Decreased	Same	14, 39
Glycine max (SBA)	Decreased	Same	14, 39
Triticum vulgaris (WGA)	Decreased	Same	14, 39
Len culinaris (LCA)	Decreased	Not evaluated	14
Ricinus communis (RCA-I)	Decreased	Not evaluated	14
Blood Group Antigens			
A and B	Decreased	Same	38
Le ^a and Le ^b	Same (negative)	Same (negative)	38
X antigen	Same (negative)	Same (negative)	38
Mucins			
Neutral Mucin	Same	Same	36
Acidic Mucin (Nonsulfated)	Elevated	Same	36

"Normal epithelium includes nodular hyperplasia.

the mean age of those with PIN (65 years) was significantly lower than those with cancer (70 years) [35].

PIN AND CANCER ARE PHENOTYPICALLY SIMILAR

There is progressive loss of markers of secretory differentiation with increasing grades of PIN, indicating progressive impairment of cell differentiation and regulatory control with advancing stages of prostatic carcinogenesis (Table 2) [13-15,36-39]. With rare exceptions, the expression of a wide variety of secretory proteins, cytoskeletal proteins, and glycoproteins is similar in PIN and invasive prostatic carcinoma, compared with normal prostatic epithelium. Nagle et al. suggested that changes in cytoskeletal proteins in PIN may affect transport of cell products, accounting for the differences in secretory protein distribution [37]. Recently, McNeal et al. have shown that reduction of cytoplasmic differentiation markers during the preinvasive phase may be followed by abrupt re-expression at the site of microinvasion [15].



Fig. 5. Frequency of PIN and cancer with increasing age in human autopsy prostates. There is a parallel increase in the frequency of PIN and cancer, according to serially sectioned autopsy prostates, although PIN appears to predate cancer by more than 10 years. (Data on PIN from [8,34]; data on cancer curve from [21].)

PIN AND CANCER ARE MORPHOMETRICALLY SIMILAR

Virtually all measures of nuclear abnormality by computer-based image analysis revealed the similarity between PIN and cancer, in contrast TABLE 3. MORPHOMETRIC FEATURES OF PROSTATIC INTRAEPITHELIAL NEOPLASIA (PIN)

Nuclear Features	PIN Compared with Normal Epithelium	PIN Compared with Cancer	References
Nuclear Area	Elevated	Same	41 45
Nuclear DNA Content (Proliferation Index)	Elevated	Same	41,45
Chromatin Heterogeneity	Elevated	Same	41
Frequency of Chromatin Clumps	Elevated	Same	41
Chromatin Condensation Level	Elevated	Same	41
Chromatin Contrast	Decreased	Same	41
Nuclear Perimeter	Elevated	Same	45
Nuclear Diameter	Elevated	Same	45
Nuclear Roundness Factor	Elevated	Same	45
Nucleolar Features			
Nucleolar Diameter	Elevated	Same	40,46,47
Multiple Nucleoli	Elevated	Same	40,47
Nucleolar Eccentricity	Elevated	Same	40,47
Nucleolar Organizer Regions (AgNORs)	Elevated	Decreased	44,46

Includes hyperplastic epithelium

with normal and hyperplastic epithelium (Table 3) [40-44]. These changes include nuclear area, DNA content, chromatin content and distribution, nuclear perimeter, nuclear diameter, and nuclear roundness. Also, most measures of nucleolar abnormality revealed the similarity between PIN and cancer, in contrast with normal epithelium [41,43,45–48]. Layfield and Goldstein found that core and needle biopsies were more reliable than fine needle aspiration in separating PIN from cancer based on a morphometric study of 50 "atypical" cases [49]. These cumulative data indicate that the morphologic continuum from PIN to cancer is characterized by progressive morphometric derangements of nuclei and nucleoli.

PIN IS FREQUENTLY ANEUPLOID

Morphologic derangements of nuclei and nucleoli observed during prostatic carcinogenesis are accompanied by progressive transformation of the normal diploid DNA content to nondiploid, according to computer-based static image analysis of tissue sections [41,45,50–52]. Montironi *et al.* suggested that two successive phases occur: the first occurs in hyperplastic epithelium and low grade PIN, and is characterized by DNA duplication without nuclear division which results in euploidy [diploid (2N) or tetraploidy (4N); the second occurs only in high grade PIN and cancer, and results in emergence of aneuploid elements (triploid, hyperdiploid, hypotetraploid, and aneuploid) [45]. Similar results were reported by Petein et al., who noted that the mean proliferative index and proportion of aneuploid cell nuclei in high grade PIN were similar to cancer, but differed significantly from hyperplastic epithelium and low grade PIN [41]. Amin et al. found an incidence of 32% aneuploidy in high grade PIN and 55% in carcinoma [50], somewhat lower than the results of Crissman et al. (57% and 62%, respectively) [52]. Weinberg and Weidner also noted concordance of DNA content in a small series of PIN and cancer, with the majority being diploid [51].

A study of 67 cases evaluated by flow cytometry by O'Malley *et al.* revealed that aneuploidy was rare in PIN (1.8%), and that there was significant discordance of results with cancer [53]. The discrepancy in results between this flow cytometry study and the static image analysis studies of PIN may be due to sampling or other variations in methods.

ANIMAL MODELS OF PIN

Studies to date have not determined whether PIN remains stable, regresses, or progresses, although the implication is that these lesions can progress. Evidence supporting this hypothesis of progression has been obtained in animal models [54–58]. Leav *et al.* reported induction of prostatic hyperplasia, "dysplasia," and carcinoma in the Noble rat by administration of testosterone in 17β -estradiol [54]. This report suggests that dysplasia may progress to carcinoma, and that long-term hormonal stimulation plays a significant role in the genesis of these lesions.

Recent work on the aging ACI/Seg rat reveals a high frequency of spontaneous development of prostatic carcinoma [56–58]. Histopathologic studies have found sequential steps which culminate in grossly manifest prostatic cancer, indicating a developmental process. Interestingly, substantial alteration of the testosterone:estrogen serum ratio with malignancy supports the promotive role of steroid hormones in carcinogenesis [56].

The Lobund-Wistar rat has a 10% incidence of prostatic carcinoma with metastases when raised in a germ-free environment [55]. Recently, Pollard *et al.* demonstrated prevention of the development of primary and metastatic tumors in this rat model by feeding synthetic retinoids [N-(4-hydroxyphenyl)retinamide] [59].

CLINICAL EVALUATION AND SIGNIFICANCE OF PIN

The clinical importance of recognizing PIN is based on its strong association with prostatic carcinoma. Because PIN has a high predictive value as a marker for adenocarcinoma, its identification in biopsy specimens of the prostate warrants further search for concurrent invasive carcinoma (Fig. 6). This is particularly



Fig. 6. Suggested diagnostic response to PIN and AAH.

true for high grade PIN; if these lesions are identified by the pathologist, close surveillance and follow-up appear to be indicated. In a series of 21 patients with PIN identified on prostate biopsy because of abnormal findings by digital rectal examination, 12 (57%) had carcinoma identified on second biopsy [60]; these results were confirmed in another study of 12 patients [61].

Carcinoma was observed in 39% of 104 followup aspiration biopsies of patients with an original diagnosis of PIN 3, according to Park et al., and an additional 35% had recurrent PIN [62]. Of 48 patients with clinical suspicion of cancer and negative aspiration biopsies, follow-up aspiration revealed PIN in 15 (31%) and invasive carcinoma in 8 (17%). In another study of PIN diagnosed by fine needle aspiration, 13 of 32 patients with high grade PIN developed cancer as compared with 3 of 23 with low grade PIN; the patients were followed for 18 months with re-biopsy [63]. These data underscore the strong association of PIN and adenocarcinoma and indicate that vigorous diagnostic follow-up is needed. When PIN is encountered in prostatic specimens, all tissue should be embedded and made available for examination; serial sections of suspicious foci may be useful. Basal cellspecific anti-keratin antibodies such as 35β -E12 (high molecular weight keratin) can be used to stain tissue sections for the presence of basal cells. Unfortunately, needle biopsy specimens and cytologic specimens sometimes fail to show the suspicious focus on deeper levels, compounding the diagnostic dilemma.

Biopsy remains the definitive method for detecting PIN and early invasive cancer, but non-invasive methods are being evaluated. By transrectal ultrasound, PIN has been reported as being hypoechoic and indistinguishable from carcinoma [35,64]. Transrectal ultrasounddirected biopsy allows localization of the needle and tissue being sampled. Repeat biopsy has been suggested by some authors if the first attempt is unrevealing. Serum PSA levels may be elevated in patients with PIN according to Brawer et al. [65]. If all procedures fail to identify coexistent carcinoma, close surveillance and follow-up are indicated. Follow-up is suggested at six-month intervals for two years, and thereafter at twelve-month intervals for life [3].

The pathologist must have an understanding

of the criteria for separating PIN from benign and malignant mimics, and should report the presence, severity, and extent of these lesions. Because our understanding and recognition of PIN is recent, some have suggested that only high grade PIN be reported to avoid diagnostic and therapeutic confusion, and this suggestion seems appropriate. Only through identification and reporting of premalignant lesions can further investigations be insured and periodic examinations undertaken. In difficult and borderline situations, the pathologist should communicate closely with the urologist and report as much information as possible. Most authors agree that the identification of premalignant lesions in the prostate should not influence or dictate therapeutic decisions.

PIN also offers promise as an intermediate endpoint in studies of chemoprevention of prostatic carcinoma. Recognizing the slow growth rate of prostate cancer and the considerable amount of time needed in animal and human studies for adequate follow-up, the noninvasive precursor lesion PIN would be a suitable intermediate histologic marker to indicate subsequent likelihood of cancer.

REFERENCES

- 1. Bostwick DG: Prostatic intraepithelial neoplasia (PIN). Urology 34(Suppl):16-22, 1989.
- Bostwick DG, Brawer MK: Prostatic intra-epithelial neoplasia and early invasion in prostate cancer. Cancer 59:788-794, 1987.
- Bostwick DG: The pathology of early prostate cancer. CA: Cancer J Clin 39:376–393, 1989.
- Bostwick DG: Premalignant lesions of the prostate. Semin Diagn Pathol 5:240-253, 1988.
- Bostwick DG, Srigley J: Premalignant lesions. In Bostwick DG (ed): "Pathology of the Prostate." New York: Churchill-Livingstone, 1990, pp 37–59.
- Boone CW, Kelloff GJ, Steele V: Histopathology of the premalignant process. Cancer Bull 43:481–484, 1991.
- Drago JR, Mostofi FK, Lee F: Introductory remarks and workshop summary. Urology 34(Suppl):2-3, 1989.
- McNeal JE, Bostwick DG: Intraductal dysplasia: A premalignant lesion of the prostate. Hum Pathol 17:64-71, 1986.
- Kovi J, Mostofi FK, Heshmat MY, Enterline JP: Large acinar atypical hyperplasia and carcinoma of the prostate. Cancer 61:555-561, 1988.
- 10. Srigley JR: Small-acinar patterns in the prostate gland with emphasis on atypical adenomatous

hyperplasia and small-acinar carcinoma. Semin Diagn Pathol 5:254–272, 1988.

- Mostofi FK: Precancerous lesions of the prostate. In Carter RL (ed): "Precancerous States." New York: Oxford University Press, 1984, pp 304-316.
- Miller A, Seljelid R: Cellular atypia in the prostate. Scand J Urol Nephrol 5:17-21, 1971.
- McNeal JE, Alroy J, Leav I, Redwine EA, Freiha FS, Stamey TA: Immunohistochemical evidence for impaired cell differentiation in the premalignant phase of prostate carcinogenesis. Am J Clin Pathol 90:23-32, 1988.
- McNeal JE, Leav I, Alroy J, Skutelsky E: Differential lectin staining of central and peripheral zones of the prostate and alterations in dysplasia. Am J Clin Pathol 89:41-48, 1988.
- McNeal JE, Villers A, Redwine EA, Freiha FS, Stamey TA: Microcarcinoma in the prostate: Its association with duct-acinar dysplasia. Hum Pathol 22:644-652, 1991.
- Amin MB, Dundore P, Schultz DS, Marsh W, Bostwick D: Architectural spectrum of high grade prostatic intraepithelial neoplasia. Mod Pathol 5:50A (abstract), 1992.
- Kovi J, Jackson MA, Heshmat MY: Ductal spread in prostatic carcinoma. Cancer 56:1566–1573, 1985.
- Becich MJ: In situ spread (ISS) on prostate needle biopsy (PNB) is predictive of invasive adenocarcinoma. Mod Pathol 5:50A (abstract), 1992.
- McNeal JE, Bostwick DG: Anatomy of the prostate: Implications for disease. In Bostwick DG (ed): "Pathology of the Prostate." New York: Churchill-Livingstone, 1990, pp 1–14.
- Srigley J, Toth P, Hartwick RWJ: Atypical histologic patterns in cases of benign prostatic hyperplasia. Lab Invest 60:90A (abstract), 1989.
- Bostwick DG, Cooner WH, Denis L, Jones GW, Scardino PT, Murphy GP: The association of benign prostatic hyperplasia (BPH) and cancer of the prostate. Cancer 7D(Suppl 1):291-301, 1992.
- 22. Greene DR, Wheeler TM, Egawa S, Dunn JK, Scardino PT: A comparison of the morphological features of cancer arising in the transition zone and in the peripheral zone of the prostate. J Urol, 1992 (in press).
- Schultz DS, Amin MB, Zarbo RJ: Type IV collagen basement membrane component of prostatic intraepithelial neoplasia and associated carcinoma. Mod Pathol 5:59A (abstract), 1992.
- Barsky SH, Siegal GP, Jannotta F, Liotta LA: Loss of basement membrane components by invasive tumors but not by their benign counterparts. Lab Invest 49:140-147, 1983.
- Beckman WC Jr, Camps JL Jr, Weissman RM, Kaufman SL, Sanofsky SJ, Reddick RL, Siegal GP: The epithelial origin of a stromal cell population in adenocarcinoma of the rat prostate. Am J Pathol 128:555-565, 1987.
- 26. Boag AH, Young ID: Type IV collagenase expression in prostatic adenocarcinoma. Mod Pathol 5:51A

(abstract), 1992.

- D'Errico A, Garbisa S, Liotta LA, Castronovo V, Stetler-Stevenson WG, Grigioni WF: Augmentation of type IV collagenase, laminin receptor, and Ki-67 proliferation antigen associated with human colon, gastric, and breast carcinoma progression. Mod Pathol 4:239-246, 1946.
- Kovi J, Mostofi FK: Atypical hyperplasia of prostate. Urology 24(Suppl):23-27, 1989.
- Troncosco P, Babaian RJ, Ro JY, Grignon DJ, von Eschenbach AC, Ayala AC: Prostatic intraepithelial neoplasia and invasive prostatic adenocarcinoma in cystoprostatectomy specimens. Urology 24(Suppl): 52-56, 1989.
- Tsukamoto T, Kumamoto Y, Masumori N, Miyao N: Studies on incidental carcinoma of the prostate. Nippon Hinyokika Gakkai Zasshi 81:1343-1350, 1990.
- Sentinelli S, Rondanelli E: La neoplasia intraepiteliale prostatica: Una nova lesione displastica della prostata. Pathologica 81:127-137, 1989.
- Kastendieck H, Helpap B: Prostatic "dysplasia/atypical hyperplasia." Urology 24(Suppl):28-42, 1989.
- Humphrey PA, Frazier HA, Paulson DF, Vollmer RT: Extent of severe dysplasia in the prostate is inversely related to pathologic stage. Mod Pathol 5:54A (abstract), 1992.
- Sakr WA, Haas GP, Cassin BJ, Pontes JE, Crissman JD: Prevalence of prostatic carcinoma in young males. An autopsy study of age and race distribution. Mod Pathol 5:58A (abstract), 1992.
- Lee F, Torp-Pedersen ST, Carroll JT, Siders DB, Christensen-Day C, Mitchell AE: Use of transrectal ultrasound and prostate-specific antigen in diagnosis of prostatic intraepithelial neoplasia. Urology 24 (Suppl):4-8, 1989.
- Humphrey PA: Mucin in severe dysplasia in the prostate. Surg Pathol 4:137-143, 1991.
- Nagle RB, Brawer MK, Kittelson J, Clark V: Phenotypic relationships of prostatic intraepithelial neoplasia to invasive prostatic carcinoma. Am J Pathol 138:119-128, 1991.
- Perlman EJ, Epstein JI: Blood group antigen expression in dysplasia and adenocarcinoma of the prostate. Am J Surg Pathol 14:810-818, 1990.
- Doria M, Jin J, Wang H, Martinez R, Matthews J, Bostwick D: Glycoconjugate expression in hyperplastic, dysplastic, and malignant prostatic epithelium. Lab Invest 4:45A (abstract), 1991.
- 40. Montironi R, Braccischi A, Matera G, Scarpelli M, Pisani E: Quantitation of the prostatic intra-epithelial neoplasia. Analysis of the nuclear size, number and location. Pathol Res Pract 187:307-314, 1991.
- 41. Petein M, Michel P, Van Velthoven R, Pasteels J, Brawer MK, Davis JR, Nagle RB, Kiss R: Morphonuclear relationship between prostatic intraepithelial neoplasia and cancers as assessed by digital cell image analysis. Am J Clin Pathol 96:628-634, 1991.
- 42. Sakr WA, Haas GP, Drozdowicz SM, Crissman JD:

Nuclear DNA content of prostatic carcinoma and intraepithelial neoplasia (PIN) in young males. An image analysis study. Mod Pathol 5:58A (abstract), 1992.

- 43. Sarkar F, Sakr W, Drozdowicz S, Sreepathi P, Crissman J: Measurement of cellular proliferation in human prostate by AgNOR, PCNA, and SPF. Mod Pathol 5:59A (abstract), 1992.
- 44. Sesterhen IA, Becker RL, Avallone FA, Mostofi FK, Lin TE, Davis CJ: Image analysis of nucleoli and nucleolar organizer regions in prostatic hyperplasia, intraepithelial neoplasia, and prostatic carcinoma. J Urogen Pathol 1:61-74, 1991.
- Montironi R, Scarpelli M, Sisti S, Braccischi A, Gusella P, Pisani E, Alberti R, Mariuzzi GM: Quantitative analysis of prostatic intra-epithelial neoplasia on tissue sections. Anal Quant Cytol Histol 12:366– 372, 1990.
- Deschenes J, Weidner N: Nucleolar organizer regions (NOR) in hyperplastic and neoplastic prostate disease. Am J Surg Pathol 14:1148-1155, 1990.
- 47. Helpap B: Observations on the number, size and location of nucleoli in hyperplastic and neoplastic prostatic disease. Histopathology 13:203-211, 1988.
- Min KW, Jin J-K, Blank J, Hemstreet G: AgNOR in the human prostatic gland. Am J Clin Pathol 95:508, 1990.
- Layfield LJ, Goldstein NS: Morphometric analysis of borderline atypia in prostatic aspiration biopsy specimen. Anal Quant Cytol Histol 13:288–292, 1991.
- Amin MB, Schultz DS, Zarbo RJ, Kubus J, Shaheen C: Computerized static DNA ploidy analysis of prostatic intraepithelial neoplasia. Mod Pathol 4:43A (abstract), 1991.
- 51. Weinberg DS, Weidner N: The relationship between prostatic intraepithelial neoplasia (PIN) and invasive prostatic carcinoma studied by static DNA cytometry. Mod Pathol 4:53A (abstract), 1991.
- Crissman JD, Sakr WA, Pontes JE: DNA quantitation in PIN and invasive carcinoma of the prostate. Mod Pathol 5:51A (abstract), 1992.
- O'Malley F, Grignon D, Keeney M, Kerkvliet N, McLean C: DNA flow cytometric studies of prostatic intraepithelial neoplasia. Mod Pathol 4:50A (ab-

stract), 1991.

- Leav I, Ho SM, Ofner P, Merk FB, Kwan PW, Damassa D: Biochemical alterations in sex hormoneinduced hyperplasia and dysplasia of the dorsolateral prostates of Noble rats. J Natl Cancer Inst 80:1045– 1053, 1988.
- Pollard M: Spontaneous prostate adenocarcinoma in aged germ-free Wistar rats. J Natl Cancer Inst 51:1235, 1973.
- 56. Isaacs JT: The aging ACI/Seg versus male Copenhagen rat as a model system for the study of prostatic carcinogenesis. Cancer Res 44:1, 1984.
- 57. Ward JM, Reznick G, Stinson SF, Lattuada CP, Longfellow DG, Camerson TP: Histogenesis and morphology of a naturally occurring prostatic carcinoma in the ACI/Seg HAP BR rat. Lab Invest 43:517, 1980.
- Shain SA, McCullough B, Nutchuck M, Boesel RW: Prostate carcinogenesis in the AXC rat. Oncology 34:114, 1977.
- Pollard M, Luckert PH, Sporn MB: Prevention of primary prostate cancer in Lobund-Wistar rats by N-(4-hydroxyphenyl)retinamide. Cancer Res 51:3610– 3611, 1991.
- Brawer MK, Bigler SA, Sohlberg OE, Nagle RB, Lange PH: Significance of prostatic intraepithelial neoplasia on prostate needle biopsy. Urology 38:103– 107, 1991.
- Weinstein MH, Epstein JI: Significance of high grade prostatic intraepithelial neoplasia (PIN) on needle biopsy. Mod Pathol 5:60A (abstract), 1992.
- Park C, Galang C, Johennig P, Maksem J, Tannenbaum M: Follow-up aspiration biopsies for dysplasia of the prostate. Lab Invest 60:70A (abstract), 1989.
- 63. Markham CW: Prostatic intraepithelial neoplasia: Detection and correlation with invasive cancer in fine-needle biopsy. Urology 24(Suppl):57–61, 1989.
- Shinohara K, Scardino PT, Carter SSC, Wheeler TM: Pathologic basis of the sonographic appearance of the normal and malignant prostate. Urol Clin North Am 16:675-691, 1989.
- Brawer MK, Lange PH: Prostate-specific antigen and premalignant change: Implications for early detection. CA: Cancer J Clin 39:361–375, 1989.